Evaluation of physical and mechanical properties of β-tri-calcium phosphate/poly-3-hydroxybutyrate nanocomposite scaffold for bone tissue engineering application

Sh. Shahi* and S. Karbasib,*

a. Department of Material Engineering, Shahreza Branch, Islamic Azad University, Shahreza, Isfahan, Iran,
b. Biomaterials Group, Advance Medical Technology Department, Isfahan University of Medical Sciences, Isfahan, Iran.

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Abstract. One of the major challenges facing researchers of tissue engineering is the scaffold design with desirable physical and mechanical properties for growth and proliferation of cells and tissue formation. In this paper, firstly, β-tricalcium phosphate (β-TCP) nanopowders with particle size of 50-70 mm were synthesized using a simple sol-gel route with calcium nitrate and potassium dihydrogenphosphate as calcium and phosphorus precursors, respectively. Then, the porous ceramic scaffold containing 40, 50, and 60 wt% of n/β-TCP was prepared by the polyurethane sponge replication method. The scaffolds were coated with P3HB for 30 sec and 1 min in order to increase the scaffold’s mechanical properties. XRD, XRF, SEM, TEM, and FT-IR were used in order to study the phase and element structure, morphology, particle size, and determination of functional groups, respectively. Based on the results of compressive strength and porosity tests, the most appropriate type of scaffold is 50 wt% of n/β-TCP immersed for 30 sec in P3HB with 75% porosity in 200-600 μm, with a compressive strength of 1.09 MPa and a compressive modulus of 33 MPa, which can be utilized in bone tissue engineering.

1. Introduction

Every year, almost two million patients worldwide undergo bone grafting procedure to regenerate bone injuries due to accidents and automobile accidents, birth defect, diseases, and different cancers [1,2]. Autografts and allografts are the current strategies for filling bone defects, but each has several disadvantages leading to limited structural and functional recovery [3,4]. For example, there are significant drawbacks to the use of autologous bone, such as limited access and unstable nature, hematoma, infection, increased operative time, blood loss, chronic donor site pain, and extra cost [4,5]. Allograft bone substitutes involve risks of immune rejection, or even lead to infectious disease transmission [6]. Synthetic bone filling materials can be prepared free of obstacles mentioned for autografts and allografts [7]. Synthetic bone filling materials have the aim of mimicking the physical and mechanical nature of native tissue and promoting osteoconduction for bone repair [3]. Then, the best strategy to treat bone injuries is the use of tissue engineering [8]. The primary goal of Tissue Engineering (TE) is to produce functional substitutes for damaged or lost tissues [9]. So far, various three-dimensional (3D) scaffolds have been produced, and then populated with signaling molecules and cells
to act as a temporary physical framework allowing new tissues to grow and reconstruct the damaged ones [10]. In other words, tissue function restoration needs a template, a scaffold, which will play the role of extracellular matrix in order to allow cells to attach, migrate, and duplicate tissue ingrowth until the new tissue is fully regenerated [11,12]. Bone tissue has a great potential for being restored to other body tissues [13]. Inherently, natural bone tissue is a composite of a collagenous hydrogel matrix consisting of about 33-43% apatite minerals, 32-44% organics, and 15-25% water on a volumetric basis [14,15], which is accountable for the remarkable strength and partial elasticity in bone tissue [16]. Moreover, hybrid structures, made of synthetic or naturally occurring ceramics and polymers, have taken advantage of this combination such as the strong compressive strength of ceramics as well as the flexibility of ductile polymers. This combination seems to be a promising choice, in particular for bone tissue engineering [17,18]. Therefore, the consequent strategy is to combine polymers and inorganic components, such as CaPs, to construct scaffolds that meet the requirements desired for bone tissue engineering [19]. Bone tissue engineering attempts to reconstruct and maintain the function of human bone tissues using the combination of cell biology, materials science, and engineering principles [8]. Both ceramic matrix and polymer matrix scaffolds have high porosity, bioactivity, biodegradability, and other physical and mechanical properties required for bone tissue engineering [20]. Ceramic matrix scaffolds have greater ability to bond directly to bone tissue compared to polymer matrix scaffolds. This means that ceramic matrix scaffolds are more bioactive than polymer matrix scaffolds and are, therefore, recommended for bone tissue engineering [20]. β-TCP [CaO (PO₄)₂], and HA [Ca₁₀ (PO₄)₆ (OH)₂] are similar to those found in human bone; therefore, they are the most commonly used ceramics in the biomedical fields [21,22]. Among these bioocermics, β-TCP and α-TCP have been widely used instead of apatites for hard tissue restoration due to their excellent tissue biocompatible, biodegradable, and osteoconductive characteristics [6,23]. For example, a biodegradabile and osteoconductive biocomposite made of β-TCP particles and gelatin was proposed [24]. This material was tested in vivo with good results without the need for a second surgical operation to remove the device after healing occurrence [6]. This property imparts significant advantage to β-TCP compared to other bioceramics, which are not resorbed and replaced by new-formed bone tissues [25]. Therefore, β-TCP bioceramics are extensively used in bone filling substituted in the field of oral plastic surgery and bone tissue engineering scaffolds [25]. Based on previous studies, a desired scaffold for bone engineering should comply with a number of mechanical and physiological demands:

1. Biocompatibility to provide cell attachment, differentiation, and proliferation;
2. Osteoconduction and osteoproduction (inducing powerful bone bonding);
3. Biodegradability at a rate of matching the rate of new tissue formation;
4. Mechanical capability in order to enable mechanical strength in load bearing sites prior to reconstruction of new tissue;
5. Interconnected porous structure for cell infiltration, tissue ingrowth, and vascularization [6,26,27].

Consequently, the optimal scaffold design and fabrication techniques must be able to create porous structures convenient for attaining the desired mechanical function and mass transport properties [28]. Polymer replication methods enable the scaffolds formation with uniform and adjustable porosity [29-31]. The method has been developed due to its interconnected pores that are similar to cancellous bone [11]. However, since CaP₄O₈ has a ceramic nature, they are brittle; therefore, biomedical applications of bare CaP₄O₈ scaffolds for bone tissue reconstruction have been largely limited by their poor mechanical properties such as compression strength, compression module, and toughness [32]. Hence, CaP₄O₈ scaffolds have been modified by introducing ductile polymer as a reinforcing phase in order to improve the mechanical properties and decrease their brittleness [13]. For example, collagen coated BCP ceramics were studied and the biocompatibility towards osteoblasts was found to increase upon coating with collagen [33]. This approach resulted in applications of conventional composite manufacturing techniques to the field of biomaterials, and the studies summarized showed that the proper combination of a brittle, hard and bioactive CaP₄O₈ matrix with a ductile polymer coating offers many advantages of both for clinical applications [6]. Poly-3-hydroxybutyrate (P3HB) is a partially crystalline polymer with a high melting temperature and a high degree of crystallinity, which can be applied as a coating in ceramic scaffolds [8]. Poly-3-hydroxybutyrate, as a member of the polyhydroxalkanoate family, known as PHB, has attracted much attention for a wide range of medical applications due to its biodegradation, which indicates a much longer degradation time compared to polymers of the poly(α-hydroxyacids) group [34,35]. Namely, PHB/carbonated ACP and PHBH/ACP biocomposites appeared to be well suited as slowly biodegradable bone replacement material [36]. Kang et al. [37] showed that the compressive strength of β-TCP scaffolds increased from 2.90 to 4.19 MPa using the infiltration of PLGA while retaining an interconnected porous structure with a porosity of 80%. Miao et al. [38] produced scaffolds...
from HA/TCP coated with the polymer PLGA, and the compressive strengths of the scaffolds increased up to 660 kPa by the PLGA incorporation in ceramic-based scaffolds. Foroughi et al. [20] indicated that HA scaffolds coating with poly-3-hydroxybutyrate, by and large, improved the compressive strength of the HA scaffolds. Their results exhibited porosity of 80-90% and compressive strength and modulus of 1.51 and 22 MPa, respectively. Bang et al. [19] evidenced that the compressive strength of α-TCP foam increased from 30 to 750 kPa by modification with interpenetration and coating of PCL. Therefore, based on the result of studies, it is clear that covering ceramic scaffold with polymer layers is one of the best achievements for production of bone tissue engineering scaffolds.

To the best of our knowledge, this is the first report on the fabrication of scaffolds from nano-β-TCP coated with P3HB. Nβ-TCP powder was prepared using a sol-gel route, and porous scaffolds were produced using a polymer replication method. In order to improve mechanical properties, scaffolds were dip-coated with P3HB. Moreover, the physical and mechanical properties of nanoβ-tricalcium phosphate-poly-3-hydroxybutyrate (nβ-TCP/P3HB) composite scaffolds were evaluated.

2. Materials and methods

2.1. Preparation of ceramic nβ-TCP nanopowder

Calcium nitrate tetrahydrate (Ca(NO$_2$)$_2$.4H$_2$O) (CNT) (Junseiechemicals Co., Ltd., S. Korea) and potassium dihydrogenphosphate (KH$_2$PO$_4$) (KDP) (Kanto Chemical Co., Inc., Japan) were used as starting calcium and phosphorus precursors. Figure 1 shows the flow chart of the sol-gel preparation of β-

![Figure 1. Flow chart of the sol-gel preparation of nβ-TCP.](image)

TCP. About 1 M CNT and 0.67 M KDP solutions were prepared by dissolving the crystals in double-distilled water. CNT solution was added drop-wise to the KDP solution while stirring. Ammonia (NH$_3$) (Daegu Chemicals and metals Co., Ltd., S. Korea) was added drop-wise to the above mixture resulting in white precipitation. The NH$_3$ was added till the precipitation was complete, and the final pH 10 was obtained. This precipitated solution was quickly stirred for 1 h and aged at room temperature for 48 h. The precipitate was filtered and washed repeatedly using the vacuum pump via lukewarm distilled water to remove NH$_3^+$ and NO$_3^-$ ions. The created precipitate was dried at 40 C for 24 h in a dry oven. This dried powder was calcined in air at 800°C for 30 min using an electrical furnace (Daesung electric, Korea) and a heating rate of 10°C/min [39]. Then, the powder produced was subjected to analysis by XRD, TEM, XRF, and FTIR tests.

2.2. Fabrication of ceramic slurry

Preparation of slurry stabilized with appropriate additives is significant and critical to scaffold fabrication; a variety of additives with a range of biological properties have been investigated [40]. In this study, β-TCP nanopowder with a particle size of 50-70 nm was utilized as the scaffold matrix. Then, nβ-TCP scaffold, containing various weight ratios of nano-β-tricalcium phosphate (40, 50, and 60 wt.%) was produced by a foam replication method. First, nβ-TCP powder (40, 50 and 60 wt.%) was slowly dispersed in double-distilled water to prevent agglomeration, and then the slurry was stirred at 350 rpm for 90 min to obtain a homogeneous solution. To maximize the mechanical strength, large amounts of solid material should precipitate from the homogenized solution to the polyurethane sponge. Ammonium poly-methacrylate (DARVAN® C-N, R.T. Vanderbilt Company, Norwalk, USA) up to 4 wt.% was used as dispersant to obtain a uniform distribution of solid substance in the ceramic slurry in order to improve its rheological properties. Carboxymethyl cellulose (CMC, Hangzhou Hongbo Chemical Co. Ltd, China) powder (4 wt.%) was gradually and slowly added to the solution in order to enhance slurry fluidity and prevent cracking of the struts during the subsequent heat-treatment process. The solution was stirred at 60°C until it was fully homogeneous.

2.3. Preparation of porous scaffold

A commercial polyurethane sponge (MEAY Co., Ltd, China; average pore size of 300-700 μm) was used in this research. Polyurethane sponge used as template that helped to manufacture ceramic cellular structures. The sponge was soaked into ceramic suspension and squeezed twice. Figure 2 shows the flow chart of the
polymer replication method used for the fabrication of nβ-TCP scaffold. This ensured that the ceramics coat the polyurethane walls to produce a replica of the reticulated foam material; the ceramic parts of the biomaterial infiltrated into the polymeric sponge pores [41]. The ceramic-coated polymeric template was dried for 24 hours at 80°C in an oven. Subsequently, the scaffolds were placed in a heat treatment furnace. nβ-TCP scaffold was formed in four stages:

1. Treatment at 600°C with a heating rate of 3°C/min;
2. Keeping at 600°C for 1 hour to completely burn the sponge;
3. An increase in temperature from 600 to 1260°C at a rate of 10°C/min;
4. Cooling to room temperature at a cooling rate of 10°C/min.

The cooled samples were removed from the furnace, measured, and then placed in desiccators. Figure 3 shows the diagram of heat treatment process of nβ-TCP porous scaffold. In Figure 4, the left sponge shows the polyurethane sponge prior to immersion in ceramic slurry, and the right scaffold shows the ceramic scaffold produced at 1260°C.

2.4. nβ-TCP scaffold coated with P3HB

One possible drawback of the method is the tendency to produce a hole in the center of each strut resulting from the removal of the polymer structure on heating. The presence of this hole can detrimentally affect the mechanical behavior of the scaffolds [42]. Therefore, the approach of filling the hole with a polymer leads to enhanced mechanical properties exploiting the interaction between the polymer and ceramics phases. In this research, 0.6 g of P3HB powder (Sigma-Aldrich, St Louis, MO, USA) was dissolved and heated in 10 mL of chloroform (Sigma-Aldrich, USA) for 6 hours in an oil bath with a reflux condenser at 60°C to obtain the polymer solution for coating nβ-TCP scaffolds. Next, nβ-TCP scaffolds were immersed in the polymer solution for 30 sec and 1 min. To achieve a uniformly coated surface and remove excess polymer solution, the samples were wrapped in aluminum foils and centrifuged at 500 rpm for 30 sec. Finally, the samples were placed in a vacuum oven at room temperature for 24 h.

2.5. Physical characterization

2.5.1. Thermal Gravimetric Analysis (TGA)

Thermal Gravimetric Analysis (TGA) measures thermal stability and compounding materials. In this research, a thermal gravimetric analysis device (TG/DTA, TGA 401, Sanatana Co.) was used to record the decrease of weight percentage of the polyurethane sponge versus temperature. The sample was heated at a rate of 1°C/min up to 600°C under nitrogen flow.

2.5.2. XRF analysis

XRF analysis was applied as chemical analysis and for determination of chemical composition of nβ-TCP powder. Elements with low atomic weight (H2, He and
Li) cannot be identified with XRF; however, all the other elements can be identified.

2.5.3. Transmission Electron Microscopy (TEM)
Transmission electron microscopy (TEM, Philips CM200 FEG, Netherlands) was used to elucidate the morphology and particle size of β-TCP powders.

2.5.4. X-ray diffraction analysis
X-ray diffraction methods (XRD, Philips X-Pert) were applied to gain information about the structural changes and phases of nβ-TCP powder, P3HB powder, nβ-TCP scaffolds with and without P3HB coating, CuK X-ray was used for the analysis. The scan rate was set to 1°/min; the imposed voltage and current were 40 kV and 30 mA, respectively, and the diffraction angle (2θ) varied from 10 to 90° at a rate of 0.4°/min.

2.5.5. FT-IR spectroscopy
Fourier transform infrared spectroscopy (FT-IR: 6300, JASCO, Japan) was used for studying the functional groups. Solid samples were prepared by pellet technique. This technique is based on the fact that a sample in KBr powder can be compressed under pressure with or without vacuum to fabricate transparent disks. Spectral analyses were performed using standard Nicolet and Microcal Origin software. FT-IR spectra were taken from nβ-TCP powder, P3HB powder, nβ-TCP scaffolds with and without P3HB coating.

2.5.6. Scanning Electron Microscopy (SEM)
Scanning electron microscopy (SEM, Philips XL-30, Netherlands) was applied to study the morphology of nβ-TCP scaffolds with and without P3HB coat. The samples were coated with gold under an argon atmosphere.

2.5.7. Porosity and density measurements
Liquid displacement was used to calculate the porosity and density of the scaffolds. Scaffold density provides information about the size, distribution, and permeability of pores and the presence of structural defects in sintered ceramic frameworks [13].

Owing to the hydrophobic properties of the polymer, 96% ethanol, which can pass easily through pores, was used instead of water. The mass of the ceramic sample (W) was measured, and a volume (V1) of ethanol was poured into a graduated cylinder and measured. The sample was immersed in ethanol for 5 min until it became saturated (V2). The difference between volumes (V2 - V1) represented the volume of the scaffold. The ethanol-soaked scaffold was removed from the graduated cylinder, and the remaining volume was recorded as V3. V1 - V3 represented the volume of ethanol absorbed by the scaffold [40].

The following equation calculates the density of scaffold (ρ) (Eq. (1)):

\[
ρ = \frac{W}{V_2 - V_3}
\]  

(1)

The following equation calculates the amount of open porosity of scaffold (ε):

\[
ε = \frac{V_1 - V_3}{V_2 - V_3}
\]  

(2)

2.6. Mechanical characterization
Machining and gripping the specimen is a major problem in the mechanical characterization of porous ceramic structures; conventional methods of mechanical characterization, such as tensile, biaxial, and impact tests, are usually inapplicable to biomaterial with porous frameworks [13]. Compression impact tests for porous bone and nβ-TCP specimens with cellular structure are instead applied [13,40]. Compressive strength and compressive modulus tests for nβ-TCP scaffolds with and without P3HB were measured using a compression impact tester (SANTAM-Eng. Design Co. LTD.) with a 10-KN load cell based upon guidelines set in ASTM-D5024-95a. The dimensions of each sample were 20×10×10 mm3 for the compression test. As ceramic scaffolds are fragile, the crosshead speed was set at 0.5 mm/min to avoid any damage to the ceramic structure. The elastic modulus was calculated as the slope of the primary linear portion of the stress-strain curve. The yield strength was calculated from the intersection of the two tangents on the curve around the yield point.

2.6.1. Statistical analysis for mechanical characterization
Statistical analysis was carried out by two-way ONAVA analysis test in IBM SPSS 722 software.

3. Results and discussion

3.1. XRF analyses of β-TCP nanopowder
The results of XRF analysis of nβ-TCP powder sample produced using the sol-gel method are presented in Table 1, indicating, with acceptable matching, that the weight percentages are similar to standard weight percentages of nβ-TCP. The calculations showed that \( \frac{W_{β-TCP}}{W_{P3HB}} \) value is equal to 1.16, remarkably close to theoretical value of 1.18. These results verified the production of nβ-TCP with desired weight percentages and excellent purity.

3.2. TEM analyses of β-TCP nanopowder
Figure 5 displays TEM micrographs of nβ-TCP powder. Morphology and size of nβ-TCP particle are shown in the image. The particle-size distribution of nβ-TCP was mainly in the wide range of 50-70 nm with irregular particle shapes.
Table 1. Chemical analysis of nano-β-TCP powder.

<table>
<thead>
<tr>
<th>Compound</th>
<th>Concentration (%W/W)</th>
</tr>
</thead>
<tbody>
<tr>
<td>CaO</td>
<td>51.1</td>
</tr>
<tr>
<td>P2O5</td>
<td>43.89</td>
</tr>
<tr>
<td>Na2O</td>
<td>1.07</td>
</tr>
<tr>
<td>SO3</td>
<td>0.972</td>
</tr>
<tr>
<td>MgO</td>
<td>0.852</td>
</tr>
<tr>
<td>SiO2</td>
<td>0.625</td>
</tr>
<tr>
<td>Cl</td>
<td>0.416</td>
</tr>
<tr>
<td>H2O</td>
<td>0.399</td>
</tr>
<tr>
<td>Al2O3</td>
<td>0.18</td>
</tr>
<tr>
<td>SrO</td>
<td>0.149</td>
</tr>
<tr>
<td>K2O</td>
<td>0.18</td>
</tr>
<tr>
<td>Fe2O3</td>
<td>0.069</td>
</tr>
<tr>
<td>CuO</td>
<td>0.037</td>
</tr>
<tr>
<td>ZnO</td>
<td>0.033</td>
</tr>
<tr>
<td>TiO2</td>
<td>0.027</td>
</tr>
<tr>
<td>LOI*</td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>100</td>
</tr>
</tbody>
</table>

*LOI: Loss On Ignition (1400°C, 90 min).

Figure 5. TEM of nano-β-TCP powder.

3.3. XRD and FTIR analyses of β-TCP nanopowder and nβ-TCP scaffold without P3HB

3.3.1. X-ray diffraction analysis

Figure 6(a) shows X-ray diffraction spectra that nβ-TCP powder produced by the route of Sanosh et al. [39]. According to XRD spectra, it is obvious that the obtained product was nβ-TCP with high purity, and no additional stable phase was revealed. Diffraction peaks for the obtained product favorably corresponded to the standard JCPDS: 9-169 [43]. β-TCP crystallizes in the rhombohedral space group R3c with unit cell parameters a = 10.439 Å, c = 37.375 Å [44]. β-TCP crystal size at 800°C was estimated up to 38.50 nm by modified Scherer equation (Monsi et al.) [45]. It should be noted that besides

![Figure 6](image-url)  

Figure 6. Pattern of XRD analysis: (a) nβ-TCP powder; and (b) nβ-TCP scaffold sintered at 1260°C.

finite crystallite size, deviations from ideal crystallinity has also a significant influence on revealing XRD peak broadening [39]. During calcination of nβ-TCP at 800°C, crystallinity increases to more than 90% due to its predominating factor in sharpening of the diffraction peaks rather than particle size; therefore, no overlaps appeared [39]. Then, by forming a solution and constructing scaffold at 1260°C, peak XRDs are shown in Figure 6(b). XRD peaks of diffraction pattern agree appropriately with those standards of β-TCP phase in the diffraction file (Card No. 00-0169) as the primary crystalline phase and with a minor quantity of other phase as the secondary phase [43]. On the other hand, as evidenced by Gao et al. [43], β-TCP started being transformed to α-TCP above 1180°C. Hence, it is not surprising that the peaks associated with the conversion of β-TCP phase into α-TCP phase appeared during sintering of β-TCP scaffolds at temperature up to 1260°C. Therefore, the diffraction peaks at 28.30°, 31.24°, and 34.60° angles related to α-TCP crystalline phase overlapped with several peaks of β-TCP crystalline phase, and acceptable matching was observed with the standard JCPDS: 9-348 [43]. Furthermore, according to comparison of Figure 6(a) and (b) at a defined angle of 47° concerning single-phase β-TCP at both patterns, β-TCP phase’s quantitative values of both patterns related to powders and sintered scaffolds were determined by using the ratio of slopes method [46]. Based on the result of quantitative phase analysis, the quantitative value of β-TCP phase decreased. This indicates the fact that β-TCP phase has been converted into α-TCP phase during sintering of β-TCP porous scaffolds above the transformation temperature of 1120°C.

3.3.2. FT-IR spectroscopy

The patterns of FT-IR β-TCP powder and β-TCP ceramic porous scaffold are shown in Figure 7(a) and (b), respectively. Figure 7(a) shows FTIR diagram related to β-TCP powder, in which strong peaks at 1092 and 1040 cm⁻¹ are typically related to the

![Figure 7](image-url)
components of triply degenerate anti-symmetric P-O stretching \( \nu_3 \) vibration. Peripheral absorption band at 962 cm\(^{-1} \) is related to \( \nu_1 \), the non-degenerate P-O symmetric vibration. 601 and 571 cm\(^{-1} \) bands in the spectra of \( \beta \)-TCP powder are related to components of triply degenerate \( \nu_3 \)O-P-O bending vibration. The peaks visible at 1640 and 3400 cm\(^{-1} \) are also attributed to the molecular and absorbed water. The clear absorption bands at 632 cm\(^{-1} \) are attributed to the stretching and vibration of the significant concentration of hydroxyl group remaining in structure [39]. A weak peak emerged near 875 cm\(^{-1} \) is attributed to the stretching vibration of P-O (H) band in HP\( \text{O}_2 \) groups. The short peaks observed near 1490 and 1426 cm\(^{-1} \) are due to \( \nu_3 \) vibration mode of CO\( \text{O}^2 \) groups. The defined bands from inorganic carbonate ion indicate that carbon of furnace atmosphere gets dissolved in the organic components and does not pyrolyze completely and may instead dissolve into \( \beta \)-TCP crystal [39]. The mineral phase of bone consists of carbonates [47]; hence, \( \beta \)-TCP may tend to have better bioactivity as a consequence of CO\( \text{O}^2 \) presence. The other peaks detected in Figure 7(b) reveal the same process mentioned in Figure 7(a).

### 3.4. XRD and FTIR analyses of \( \beta \)-TCP scaffold with and without P3HB

#### 3.4.1. Thermal Gravimetric Analysis (TGA) of polymer foam

In order to prevent cracks in the ceramic structure, there should be sufficient time for burning polymeric foams. Figure 8 shows the weight change of the polyurethane foam relative to temperature. TGA curve demonstrated that the polymer sponge would undergo weight decline in several stages, shown in the figure. The main part of the polymer was removed at temperatures between 350°C and 600°C; therefore, it is necessary to take into account the slow heating rate at this stage to prevent structural damage caused by emissions from thermal degradation of polymers. At temperatures above 600°C, the polymer foam had a low weight decrease due to the amount of carbon produced from the decomposition of the polymer process dioxide. Complete combustion of polymeric foams occurred at 600°C.

#### 3.4.2. X-ray diffraction analysis

Figure 9(a), (b), and (c) show X-ray diffraction patterns of \( \beta \)-TCP, P3HB, and \( \beta \)-TCP/P3HB scaffolds, respectively. The pattern of XRD in Figure 9(a) demonstrates the process mentioned in Section 3.1.1 about XRD of \( \beta \)-TCP porous scaffold. In Figure 9(b), the main peaks of P3HB are detected, and Figure 9(c) is attributed to XRD pattern of \( \beta \)-TCP scaffold coated with P3HB; according to the standards of JCPDS: 009-0169 and JCPDS: 009-0348, the peaks of 13.86, 17.30, and 21.88 angles appeared in Figure 9(c) are assigned to P3HB. Since two of the strong peaks of P3HB are observed in Figure 9(c), it can be evidenced that the composite structure was a hybrid scaffold of \( \beta \)-TCP/P3HB.

#### 3.4.3. FT-IR spectroscopy

The patterns of FTIR \( \beta \)-TCP scaffold and P3HB pure polymer and \( \beta \)-TCP/P3HB scaffolds are shown in Figure 10(a), (b), and (c), respectively. The pat-
Figure 10. Pattern of FT-IR analysis: (a) nβ-TCP scaffold, (b) P3HB, and (c) nβ-TCP/P3HB scaffold.

Figure 11. Pattern of FT-IR analysis: (a) nβ-TCP and nβ-TCP/P3HB scaffolds and (b) P3HB polymer and nβ-TCP/P3HB scaffold.

TCP/P3HB nanocomposite is shifted to a lower wave number up to 1721 cm$^{-1}$, which possibly indicates hydrogen bond formation between P3HB carbonyl groups and OH groups of β-TCP particles. As previously mentioned, the twin peak observed in range of between 2874 and 2972 cm$^{-1}$ is assigned to stretching aliphatic band of C-H in pure polymer in which the same absorption band in the composite scaffold is observed with lower intensity in narrow range of wave number. The connection between the scaffold ceramic background and polymer molecules was confirmed by FTIR spectra [48].

3.5. SEM of nβ-TCP scaffold with and without P3HB

Figure 12(a) depicts the electron microscopic images of polyurethane foam with open pores in range of 300-700 μm before immersion in the ceramic slurry. Figure 12(b) exposes porous nβ-TCP scaffold with pore diameters ranging from 200 to 600 μm. Figure 12(c) displays the micropores fabricated from nβ-TCP particles; these micropores promote tissue growth [20]. Figure 12(d) shows porous nβ-TCP scaffold immersed in the polymer for 30 sec, and Figure 12(e) illustrates nβ-TCP scaffold immersed in the polymer for 60 sec. As observed in Figure 12, SEM images show uniform porosity in size 200 to 600 μm, which are appropriate for adhesion and migration of osteoblast cells [49].
Miao et al. [38] reported HA/TCP composite scaffold porous structure with open macropores ranging from 300 to 700 μm in diameter by the scanning electron micrograph observations. Also, Murphy et al. [49] evidenced the effect of scaffold pore size on cell attachment, induction, differentiation, proliferation, and migration in bone tissue engineering, and discovered that 325 μm mean pore size is appropriate for bone tissue engineering scaffolds. Regarding Figure 12(d) and (e), it can be proved from these images that adding P3HB to nβ-TCP ceramic scaffold and also the immersion time increase of scaffolds in the polymer decrease the porosity of scaffolds while maintaining a more interconnected porous structure which is beneficial for cells adhesion. Fabrication of pore interconnected porous structure is needed for the precise diffusion of nutrients and gases, such as oxygen, and for the removal of metabolic wastes resulted from the activity of migrated cells that had, meanwhile, grown into the porous scaffolds, thereby mimicking the cancellous bone structure [11, 50]. Therefore, it is concluded that adding P3HB to nβ-TCP ceramic scaffold decreases the pore size of scaffolds, but it does not have harmful effect on them, because the scaffolds coated with a polymer have acceptable pore sizes for growing of bone cells, making them suitable candidates for bone replacement.

3.6. Porosity measurements
The porosity measurement was performed on nβ-TCP scaffolds with and without P3HB coating in 30 and 60 sec, and the porosity range was 70-84%. The results of porosity percentages of the samples are shown in
Table 2. Mean porosity of nβ-TCP scaffold with and without P3HB.

<table>
<thead>
<tr>
<th>Sample</th>
<th>Porosity % without P3HB</th>
<th>Coated P3HB in 30 sec</th>
<th>Coated P3HB in 1 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>S_1*</td>
<td>84 ± 1</td>
<td>77 ± 1</td>
<td>74 ± 1</td>
</tr>
<tr>
<td>S_2</td>
<td>80 ± 1</td>
<td>75 ± 2</td>
<td>73 ± 1</td>
</tr>
<tr>
<td>S_3</td>
<td>75 ± 2</td>
<td>72 ± 1</td>
<td>70 ± 1</td>
</tr>
</tbody>
</table>

* S_1: 40% β-TCP scaffold; S_2: 50% β-TCP scaffold; S_3: 60% β-TCP scaffold

Table 2. The results showed that with the increase of weight percentage of β-TCP, the porosity percentage was reduced. It is also concluded that adding P3HB to nβ-TCP scaffolds decreased their porosity; this is despite the fact that it did not have adverse effect on the scaffolds, and it allowed the preservation of pores interconnection after infiltration process. According to the results of porosity measurements, it may be concluded that the samples immersed for 30 sec in P3HB had higher porosity than the samples immersed for 1 min in P3HB. Mondal et al. [28] revealed that the porosity of Labeco rohitu fish scale-derived scaffold was about 30-33%. Kang et al. [37] determined that the porosity of β-TCP scaffold was about 82.49% and slightly declined up to 80.65% after PLGA infiltration. In a similar study, Chen et al. [51] demonstrated an about 2.2% devaluation in bio-glass macroporosity due to PDLA polymer coating. In addition, Miao et al. [38] proved that the total porosity of HA/TCP scaffolds was estimated to be up to 87% and indicated ~2% macroporosity reduction due to coating and interpenetrating with the PLGA polymer. Furthermore, according to previous studies of Miao et al. [52,53], the removal of crack-like defects, owing to PLGA polymer phase filling into the open voids, was demonstrated. The crack-like defects of uncoated scaffolds caused the burning off of the PU sponge, initially centered with the ceramic particles [38]. Natural reduction of mechanical properties is brought about by the flaws such as cracks. Therefore, when the scaffolds after the polymer solution dipping were placed into centrifuge, the centrifugal effect caused the polymer to fill the crack-like defects [38].

3.7. Compressive strength and compressive modulus of nβ-TCP scaffolds with and without P3HB

Figure 13(a) shows representative stress-strain curve obtained in uni-axial tests performed on bare nβ-TCP scaffolds with various weight percentages of β-TCP ceramic. This figure illustrated that β-TCP failed in an elastic response of a brittle ceramic (i.e., within the linear region of the curve) obtained by disjointed rods that would collapse presumably by brittle fracture domination of struts [37]. The onset of the first of these cracks occurred during the elastic regime (i.e., within the linear region of the curve) and was usually related to the maximum applied load, thereby determining the compressive strength of the scaffold [54]. It should be noted that compressive modulus of scaffolds is achieved by steep calculation of strain-stress curve in elastic regime. Based on the results, on one hand, compressive strength and compressive modulus of nβ-TCP scaffolds increased by the weight percentage increase of β-TCP; on the other hand, the porosity percentage decreased. The results of porosity percentage, compressive strength, and compressive modulus of uncoated and coated β-TCP scaffolds with various weight percentages are presented in Tables 2, 3, and 4, respectively. The compressive strength of a scaffold depends on its macroporosity, macro-pore size, macro-pore geometry, and the strength of the struts [38]. The compressive
strength test was also performed on nβ-TCP scaffold with and without P3HB. The compression curves of
bare 50% β-TCP scaffold and PHB-coated 50% β-TCP scaffolds for 30 sec and 1 min were exhibited in Figure
13(b). The fracture phenomena also occurred in the coated scaffolds, but the curves indicated a significant
increase of this linear elastic region range; moreover, the yield point substantially moved to a higher point,
and this proves the improvement of strengthening after coating by polymer which is superior when compared
with pure β-TCP scaffolds [37]. As seen in Tables 2 to 4, the results achieved in the present study clearly
showed that the mechanical performance of β-TCP scaffold can be enhanced remarkably by the incorpora-
tion of ductile polymer. In addition, they showed that with increasing immersion time of the polymer,
the strength and modulus increased; by contrast, the porosity percentage decreased. This might explain that
the mechanical behaviors of β-TCP scaffold improved by the substantial adhesion between β-TCP particles
and PHB [19]. According to the achievements of Nalla et al. [55], the main toughening mechanism in bone was
the crack bridging by the collagen polymer fibrils. The fracture surface showed the formation of fibrils which is
related to that of collagen fibrils in cortical bone [55]. Therefore, it could be concluded that PHB polymer
infiltration not only improved the mechanical strength of β-TCP ceramic scaffold, but also toughed β-TCP
scaffold. Kang et al. [37] produced scaffolds from β-TCP coated with a PLGA polymer. The compressive
strength of the scaffold before coating was 2.90 MPa that increased to 1.44 MPa by the interpenetration
of PLGA. Miao et al. [38] also showed that by PLGA infiltration in HA/TCP, hybrid scaffolds increased their
compressive strength up to 600 KPa. Bang et al. [19] exhibited that the compressive strength of α-TCP foam
increased from 30 ± 10 kPa to 750 ± 20 kPa when it
was immersed in PCL polymer solution. Finally, to
choose the most appropriate scaffold and compare it
with regard to the obtained results from porosity per-
centage, compressive strength, and modulus, Table 5
was prepared. Based on the data available in these
tables, the most proper scaffold for bioactivity and
biodegradability tests can be selected. It should be
noted that the suitable scaffold to use in tissue engi-
neering and in studying bioactive and biodegradable
behaviors should have porosity percent in range of 75-
80%, requiring compressive strength and modulus [56].
As seen in Table 5, β-tricalcium phosphate 40 wt.%
scaffold has suitable porosity percent; by contrast, it has
low compressive strength and modulus. Beta-
tricalcium phosphate 60 wt.% scaffold has low porosity
percent; by contrast, its compressive strength and
modulus are high. Hence, according to the results of
porosity percentage, compressive strength, and modu-
lus shown in Table 5, the most appropriate specimen
for bioactive and biodegradable tests is β-tricalcium
phosphate 50 wt.% scaffold without and with PHB
coating for 30 sec. The data of porosity, compressive
and strength modulus for 50 wt.% β-TCP scaffold are
revealed and identified with highlights in this table.

3.7.1. Statistical analysis for mechanical
characterization
As previously mentioned, results of compressive strength values related to nβ-TCP scaffold without and
with P3HB for various weight percentages are shown in Table 3. According to statistical analysis, P-values
for Scaffold without coating and scaffold with P3HB for 30 and 60 sec are less than 0.05, and the assumed lack
of difference is rejected. It means that, statistically, there is a significant difference between compressive
strength of scaffold without coating and scaffold with
P3HB for 30 and 60 sec. Furthermore, Since P-values

| Table 3. Mean compressive strength of nβ-TCP scaffolds with and without P3HB (P < 0.05, n = 3). |
| Sample | Without coat | Coated P3HB in 30 sec (MPa) | Coated P3HB in 1 min (MPa) |
| S1* | 0.1727 ± 0.0892 | 0.6335 ± 0.0460 | 0.9872 ± 0.0992 |
| S2 | 0.5657 ± 0.3524 | 1.0901 ± 0.21 | 1.1419 ± 0.4872 |
| S3 | 1.0939 ± 0.3691 | 1.4276 ± 0.2153 | 1.59 ± 0.6788 |

* S1: 40% nβ-TCP scaffold; S2: 50% nβ-TCP scaffold; S3: 60% nβ-TCP scaffold.

| Table 4. Mean compressive modulus of nβ-TCP scaffolds with and without P3HB (P < 0.05, n = 3). |
| Sample | Without coat | Coated P3HB in 30 sec (MPa) | Coated P3HB in 1 min (MPa) |
| S1* | 16.13 | 20.06 | 27.603 |
| S2 | 17.234 | 33.11 | 35.2 |
| S3 | 17.7 | 49.593 | 63.51 |

* S1: 40% nβ-TCP scaffold; S2: 50% nβ-TCP scaffold; S3: 60% nβ-TCP scaffold.
Table 5. Porosity, compressive strength, and modulus of ceramic scaffold for 40, 50, and 60 wt.% of nβ-TCP with and without P3HB coating.

<table>
<thead>
<tr>
<th>Type of scaffold</th>
<th>nβ-TCP %</th>
<th>Porosity %</th>
<th>Compressive strength (MPa)</th>
<th>Compressive modulus (MPa)</th>
</tr>
</thead>
<tbody>
<tr>
<td>β-Tricalcium phosphate</td>
<td>40</td>
<td>83.90</td>
<td>0.1727</td>
<td>16.13</td>
</tr>
<tr>
<td></td>
<td>50</td>
<td>80.49</td>
<td>0.5657</td>
<td>172334</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>74.56</td>
<td>1.0593</td>
<td>17.7</td>
</tr>
<tr>
<td>With 30 sec coating</td>
<td>40</td>
<td>76.84</td>
<td>0.6335</td>
<td>20.06</td>
</tr>
<tr>
<td></td>
<td>50</td>
<td>75.31</td>
<td>0.9019</td>
<td>33.11</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>71.80</td>
<td>1.4276</td>
<td>43.503</td>
</tr>
<tr>
<td>With 60 sec coating</td>
<td>40</td>
<td>74</td>
<td>0.9872</td>
<td>27.603</td>
</tr>
<tr>
<td></td>
<td>50</td>
<td>73.24</td>
<td>1.1419</td>
<td>35.2</td>
</tr>
<tr>
<td></td>
<td>60</td>
<td>70</td>
<td>1.59</td>
<td>63.51</td>
</tr>
</tbody>
</table>

for Scaffold with various weight percentages are less than 0.05, the assumed lack of difference is rejected. It means that, statistically, there is a significant difference between compressive strengths of 40%, 50%, and 60% β-TCP scaffolds. In fact, confidence coefficient of this mechanical test is 95%.

Similarly, the results of compressive modulus related to nβ-TCP scaffold without and with P3HB for various weight percentages are shown in Table 4. Since P-values for scaffold without coating and scaffold with P3HB coating for 30 and 60 sec are less than 0.05, the assumed lack of difference is rejected. It means that, statistically, there is a significant difference between compressive modulus of scaffold without coating and scaffold with P3HB for 30 and 60 sec. In addition, since P-values for 40, 50, and 60 wt.% scaffolds are less than 0.05, the assumed lack of difference is rejected. It means that, statistically, there is a significant difference between compressive modulus of 40% β-TCP, 50% β-TCP, and 60% nβ-TCP scaffold. In fact, confidence coefficient of this mechanical test is 95%.

4. Conclusion

In this research, β-TCP ceramic scaffold was constructed using a polymeric sponge method, and it was coated with P3HB. The results of FTIR, XRF, and XRD analyses confirmed the purity of fabricated nβ-TCP powder by using the sol-gel method, and the results of TEM analysis emphasized that the particle size of nβ-TCP is about 50-70 nm and has irregular morphology. The findings of XRD and FTIR of scaffolds also proved the production of nβ-TCP/P3HB composite scaffold. Porosity is a significant factor to design a scaffold. In fact, the production of scaffold with designed pore architecture is an essential request for bone tissue engineering. The results of SEM analysis and porosimetry indicated that adding P3HB to nβ-TCP scaffold resulted in a decrease in the amount of porosity, but the increase of size, volume, and interconnectivity of porosity was also maintained. The results of mechanical test showed that the increase of the weight percentage of β-TCP and adding P3HB to nβ-TCP scaffold brought about an increase in compressive strength and compressive modulus. In the end, according to the comparison of result collection of mechanical and porosimetry tests, it is evidenced that the optimal specimen was scaffold with 50 wt.% of nβ-TCP and coating time of 30 sec which has 1.09 MPa compressive strength, 33.11 MPa compressive modulus, and 75% porosity. Therefore, this scaffold has a promising possibility to be applied as a bone filling substitution in tissue engineering field.

References


Biographies

Shabnam Shahi received her BS degree in Ceramic-Material Engineering from Shahreza Islamic Azad University, Iran, in 2011, and MS degree in Ceramic-Material Engineering from Shahreza Islamic Azad University, Iran, in 2015. She is currently pursuing research in tissue engineering.

Saeed Karbasi received his BS degree in Material Engineering from Shiraz University, Iran, in 1994 and MS and PhD degrees in Biomedical Engineering from Amirkabir University, Iran, in 1997 and 2005, respectively. He is currently an Associate Professor at Isfahan University of Medical Sciences, Iran. His research interests include biomaterials, tissue engineering, and nanobiomaterials.