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Sulfated polysaccharide coated $BaFe_{12}O_{19}$: A magnetically separable bifunctional catalyst for the synthesis of benzopyranopyrimidines derivatives and its antibacterial activity evaluation

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KEYWORDS

Marine sulfated polysaccharide; Barium hexaferrite; Magnetic nanoparticle; Benzopyranopyrimidines; Heterogeneous catalyst; Green chemistry. Abstract. Marine sulfated polysaccharide Irish Moss (IM) coated $BaFe_{12}O_{19}$ nanocomposites were synthesized and characterized by a Fourier Transform Infrared Spectrometer (FT-IR), Scanning Electron Microscope (SEM), X-Ray Diffraction (XRD), Vibrating-Sample Magnetometer (VSM), and Thermal Gravimetric Analysis (TGA). The indisputable privilege of $BaFe_{12}O_{19}$ @IM as a recyclable acid-based bifunctional catalyst has been studied in the preparation of benzopyranopyrimidines via a pseudo-four-component reaction of salicylic aldehydes, malononitrile, and various amines. A catalytic amount of $BaFe_{12}O_{19}$ @IM has shown high catalytic activity and stability with negligible detriment in its efficiency over five catalytic cycles. The catalytic property-catalytic performance associations clearly showed the synergistic effect between Irish moss, as the major active phase, and barium ferrite nanoparticles, enabling catalyst separation in a magnetic field. Along with the catalytic activity, a study on the antibacterial performance of $BaFe_{12}O_{19}$ @IM nanocomposites possess antibacterial activity against Gram-positive Staphylococcus aureus (S. aureus).

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1. Introduction

Today, magnetic nanoparticles (MNPs) are noteworthy in the development of various heterogeneous nano catalytic systems with increased biocompatibility [1–

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4]. Among various magnetic materials reported in the literature, barium hexaferrites are the most widely used permanent magnets and still promising because of their chemical and thermal stability, low production cost, and corrosion resistivity, besides their nontoxicity [5–10]. However, bare magnetic particles tend to be readily aggregated which can be avoided by using different organic or inorganic coatings on the core/shell frame or by using synthetic polymers or natural polysaccharides as capping agents [11–19]. Natural polysaccharides IM or carrageenan moss extracted from red algae comprise nearly one sulfate monoester group per sugar unit. The

acidic characteristic of carrageenan, which is caused by its high sulfate content, accounts for many of its important properties [20–22]. It is important to mention that they are strongly anionic polymers because of their half-ester sulfate segments (Figure 1) [23]. Both sulfate and hydroxyl groups as Brønsted-Lowry basic and acidic sites render IM an effective catalytic for organic reactions. In addition, polymer-inorganic nanocomposites due to their biodegradability, non-toxicity, thermal and chemical stability constitute a new platform for the design of heterogeneous catalysts [24–28].

Due to the various applications of benzopyranopyrimidine scaffolds, such as cytotoxic activity against cancer cell lines [29], anti-aggregating, anti-fungal, antimicrobial activities [30,31], and their fundamental pharmacological properties, special attention has been paid to the synthesis of benzopyranopyrimidine scaffolds as an impressive class of heterocycles [32–42].

In continuation of this research, the catalytic activity of hybrid nanocomposites based on natural polysaccharides IM in several multicomponent reactions (MCRs) has been investigated [18,19]. In the present work, $BaFe_{12}O_{19}@IM$ as a novel and reusable biomagnetic heterogeneous nanocatalyst was prepared and used in the synthesis of benzopyranopyrimidine derivatives through one-pot multicomponent condensation of salicylaldehyde derivatives **1a**-g, malononitrile (2), and secondary amine 3a-e at ambient temperature (Figure 2). The present study demonstrates that the prepared nano biocatalyst have high thermal and chemical stability, ferromagnetic property, and easy renewability. On the other hand, in recent decades, antibacterial compounds containing metallic



Figure 1. Chemical structure of carrageenan moss.

nanoparticles have been the subject of growing research to counter the resistance of microorganisms to antibiotics. Therefore, the antibacterial activity of the designed nanocomposites against two bacterial strains was investigated.

2. Experimental

2.1. Materials and methods

All materials were purchased from Merck or Sigma Aldrich and used without further purification. IR spectra were recorded using KBr discs on a Shimadzu FT-IR-470 spectrophotometer. X-Ray Diffraction (XRD) measurements were carried out using a Philips analyzer, and the calcination of the catalyst was performed using an Exciton oven. Magnetic measurements were performed using VSM model MDKFD from Danesh Pajohan Kavir Co., Kashan. The FESEM images were recorded using a ZEISS instrument, SIGMA VP model, Germany. The ¹H and ¹³C spectra were recorded on a Bruker DRX 500-Avance (500 and 125 MHz, respectively) in CDCl₃, internal standard-TMS. Melting points were determined by electro thermal 9100 apparatus and are uncorrected. Sonication for synthesis of the catalyst was performed by Elma at 60 Hz. Thermogravimetric analyses (TGA) were performed by the D-32609 Hullhorst apparatus.

2.2. Synthesis of barium hexaferrite magnetic nanoparticles

Barium hexaferrite magnetic nanoparticles (BaFe₁₂ O_{19}) were prepared using a sol-gel method. For this purpose, an aqueous solution of barium ferrite was prepared by dissolving barium nitrate (Ba(NO₃)₂ (0.52 g, 1.99 mmol), and ferric nitrate.nonahydrate (Fe(NO₃)₃.9H₂O) (9.6 g, 23.76 mmol) in 100 ml of deionized water, followed by the addition of citric acid (4.5 g) under vigorous stirring. In order to achieve a stable dispersion, the mixture was sonicated for 15 min using an ultrasonic bath. The pH of the solution was then adjusted to 8.0 by adding an ammonia solution (25%), and the mixture was oven dried at 80°C. The



Figure 2. Synthesis of benzopyranopyrimidines in the presence of BaFe₁₂O₁₉@IM magnetic nanocatalyst.

product was calcinated at 750° C for 2 h to furnish the desired BaFe₁₂O₁₉ magnetic nanoparticles.

2.3. Preparation of BaFe₁₂O₁₉@IM nanocomposite

Irish moss (7.8 g) and synthetic $BaFe_{12}O_{19}$ (3.12 g, 2.8 mmol) were added to a mixture of H_2O : ETOH (15 ml, 2:1). The mixture was stirred for 2 h and thereafter sonicated for 40 min. After evaporation of the solvent, the obtained $BaFe_{12}O_{19}/IM$ was oven dried for 6h nanocomposite (Figure 3).

2.4. General procedure for synthesis of benzopyranopyrimidine derivatives

A mixture of salcilaldehyde (2 mmol), malononitrile (1 mmol), secondary amines (1 mmol) and $BaFe_{12}O_{19}$ @IM nanocomposites (15 mg) in EtOH (4 ml) was stirred at room temperature for 1 h. The reaction advancement was monitored by TLC (eluent, EtOAc/n-hexane, 1:3). At the end of the reaction, the catalyst was collected by an external magnet, and washed for the next experiment. The precipitate was washed several times with DI water and EtOH to obtain the pure product.

2.5. Spectral data of the selected products 2-(4-morpholino-5H-chromeno[2,3-d]pyrimidin-2yl)phenol(4a)

Yellow solid; mp. 197-199°C: IR (KBr): 3300, 2856, 2949, 1610, 1535, 1440, 1384, 1388, 1251, 1118, 1110, 1018, 946, 864, 823, 754 cm⁻¹. ¹HNMR (500 MHz, CDCl₃) ppm: 3.48–3.50 (t, J = 4.9 Hz, 4H), 3.90–

3.92 (t, J = 4.8 Hz, 4H), 3.91 (s, 2H), 6.90–6.93 (t, J = 7.4 Hz, 1H), 6.97–6.98 (d, J = 8.15 Hz, 1H), 7.10–7.13 (t, J = 7.1 Hz, 1H), 7.17–7.21 (t, J = 8.3 Hz, 2H), 7.24–7.27 (m, 1H), 7.34–7.37 (t, J = 8.1 Hz, 1H), 8.38–8.40 (d, J = 7.8 Hz, 1H), 13.11 (s, 1H) ; ¹³ CNMR (125 MHz, CDCl₃)TM ppm: 25.7, 48.8, 66.8, 76.9, 77.1, 77.4, 97.8, 117.2, 117.7, 119.0, 119.2, 124.6, 128.5, 128.6, 129.3, 133.1, 150.6, 160.5, 162.3. m/z: 361.14 (M+). Anal. Calcd for C₂₁H₁₉N₃O₃: C, 69.79; H, 5.30; N, 11.63; O, 13.28.

4-chloro-2-(7-chloro-4-morpholino-5Hchromeno[2,3-d] pyrimidin-2-yl)phenol (4h)

Yellow solid; mp. 249-251°C: IR (KBr): 3735, 3305, 3041, 1649, 1548, 1469, 1240, 1135, 1018, 939, 860, 819, 730, 669 cm⁻¹: ¹HNMR (500 MHz, CDCl₃) ppm: 3.47 (t, J = 5.0 Hz, 4H), 3.84 (t, J = 5.0 Hz, 4H), 3.90 (s, 4H), 6.85–6.87 (d, J = 8.7 Hz, 1H), 7.07–7.09 (d, J = 8.6 Hz, 1H), 7.17 (s, 1H), 7.19–7.21 (d, J = 8.7 Hz, 1H), 7.23–7.25 (d-d, J = 2.6 Hz, 1H), 8.26 (s, 1H), 12.97 (s, 1H); ¹³CNMR (125 MHz, CDCl₃) ppm: 25.4, 48.6, 64.0, 66.6, 97.7, 118.4, 118.6, 119.1, 119.3, 120.5, 123.8, 128.1, 128.5, 128.9, 129.6, 132.8, 148.7, 158.9, 161.0,164.8. m/z: 429.06 (M+). Anal. Calcd for C₂₁H₁₇Cl₂N₃O₃: C, 58.62; H, 3.98; Cl, 16.48; N, 9.77; O, 11.15.

5-methoxy-2-(8-methoxy-4-morpholino-5Hchromeno[2,3-d] pyrimidin-2-yl)phenol (40)

White solid; mp. 210-213°C: IR (KBr): 3500, 2900,



Figure 3. Preparation of BaFe₁₂ O₁₉@IM nanocomposite.

2846, 1590,1522, 1435, 1368, 1276, 1204, 1108,1023 , 813, 617 cm⁻¹; ¹HNMR (500 MHz, CDCl₃) ppm: 3.44 (m, 4H), 3.79 (s, 6H), 3.83 (s, 2H),3.88 (t, J = 4.4 Hz, 4H), 6.45–6.48 (m, 2H), 6.66 (d-d, J = 2.35 Hz, 1H), 6.70 (s, 1H), 7.05 (d, J = 8.4 Hz, 1H), 8.26 (d, J = 8.75 Hz, 1H), 13.29 (s, 1H); ¹³CNMR (125 MHz, CDCl₃) ppm: 24.8, 48.6, 55.3, 55.4, 66.6, 97.1, 101.2, 102.0, 106.6, 110.9, 111.1, 111.7, 129.0, 130.4, 151.0, 159.6, 161.9, 162.1, 163.6, 164.7 m/z: 421.16 (M+). Anal. Calcd for C₂₃H₂₃N₃O₅: C, 65.55; H, 5.50; N, 9.97; O, 18.98.

3. Results and discussion

The biomagnetic $BaFe_{12}O_{19}$ @IM was synthesized according to the method described in Figure 3 Referring to the synthesis strategy presented, barium ferrite nanoparticles were first synthesized, and then the IM was added to the reaction mixture and sonicated for 40 min. The resulting precipitate was then dried to obtain $BaFe_{12}O_{19}$ @IM magnetic nanocomposites. The synthesized $BaFe_{12}O_{19}$ @IM was characterized using FT-IR spectroscopy, TGA analysis, VSM analysis, SEM image, and XRD analysis.

3.1. Characterization of the nanocomposites 3.1.1. FT-IR analysis

FTIR spectra of $BaFe_{12}O_{19}$ nanoparticles, IM, and $BaFe_{12}O_{19}$ @IM nanocomposites were exposed in Figure 4. The peaks at 426.1 and 584.0 cm⁻¹ in the IR spectrum of $BaFe_{12}O_{19}$ are related to the metal-oxide stretching vibration originated from the $BaFe_{12}O_{19}$ crystalline structure. The broad absorption band at 3400 cm⁻¹ corresponds to the stretching vibrations of the O-H groups, and the absorbance peaks at 1650.7 and 1537.6 cm⁻¹ are due to the bending vibrations of hydroxyl groups. In the IM spectrum, absorption bands at ~ 1249 cm⁻¹ are attributed to S-O asymmetric stretching in the sulphate group, absorption



Figure 4. FTIR analysis of the BaFe₁₂O₁₉ nanoparticles, IM, and BaFe₁₂O₁₉@IM nanocomposite.

bands at ~ 1010 and 1065 cm⁻¹ are credited to C-O and C-OH stretching and 842.4 cm⁻¹, $\alpha(1,3)$ Dgalactose C-O-S stretching vibrations. By conferring on the BaFe₁₂O₁₉ @IM spectrum, the presence of the metal oxide absorption bands at 426.1 and 584.0 cm⁻¹ and all the absorption bands coherent with IM confirms that the chemical structure of the polysaccharide and of the nanoparticles of BaFe₁₂O₁₉ has been preserved after the heat treatments. Furthermore, it is clearly shown that its structure is preserved after recycling.

3.1.2. Thermogravimetric analysis (TGA)

The TGA analysis for IM, BaFe₁₂O₁₉ and $BaFe_{12}O_{19}$ @IM was performed to investigate their thermal stability (Figure 5). The samples were heated from ambient temperature to 600°C at a constant heating rate. As can be seen in the IM TGA curve, there is a two-step decomposition for IM. The first weight loss up to $\sim 200^{\circ}$ C is attributed to loss of adsorbed water. The subsequent weight loss occurring between 200 and 600°C, at about ~ 25%, can be attributed to the degradation of the polysaccharide. In comparison with IM, BaFe₁₂O₁₉@IM nanoparticles showed an appropriate resistance to the decomposition up to 300°C. This resistance is due to the existence of BaFe₁₂O₁₉ showing significantly thermal stability. Fast degradation that arose around 300°C to 400°C corresponds to $\sim 24\%$ weight loss due to the degradation of the polysaccharide, followed by the last mass loss and formation of carbonaceous material.

3.1.3. Vibrating Sample Magnetometer (VSM)

Figure 6 shows the magnetic hysteresis loops of $BaFe_{12}O_{19}$ nanoparticles and $BaFe_{12}O_{19}$ @IM nanocomposites, which were studied using VSM analysis in the range of -8500 < Oe < 8500 applied field. The magnetic saturation (M_s) and coercivity (H_c) of the hard-ferromagnetic barium hexaferrite were 48.60 emu/g and 4688.98 Oe, respectively. Re-



Figure 5. The thermogravimetric analysis of IM, $BaFe_2O_{19}$ and $BaFe_2O_{19}$ @IM.

Entry	Sample	$M_s~({ m emu/g})$	$M_r~(m emu/g)$	$H_{\rm c}~({\rm Oe})$
1	$\mathrm{BaFe_{12}O_{19}}$	48.60	30.64	4688.98
2	$\mathrm{BaFe_{12}O_{19}}@\mathrm{IM}$	39.93	21.34	2630.93

Table 1. Magnetic parameters of the $BaFe_{12}O_{19}$ and $BaFe_{12}O_{19}@IM$ nanostructures.



Figure 6. Magnetic hysteresis loops of the $BaFe_{12}O_{19}$ nanoparticles and $BaFe_{12}O_{19}@IM$ nanocomposite.

sults indicated that the M_s of the BaFe₁₂O₁₉ @IM nanocomposites was diminished from 48.60 emu/g to 39.93 emu/g, which was related to the mass fraction of the non-magnetic polysaccharide within the nanocomposite. BaFe₁₂O₁₉ @IM nanocomposites also show a hard-magnetic behavior referring to their broad hysteresis loops. Magnetic reusability originated from heat resistant barium hexaferrite is the most significant feature of the BaFe₁₂O₁₉ @IM nanocomposite. It should be noted that the magnetic property of the nanocomposite BaFe₁₂O₁₉ @IM was good enough for easy separation in a magnetic field. Table 1 shows the magnetic parameters of BaFe₁₂O₁₉ @IM nanoparticles and BaFe₁₂O₁₉ @IM nanocomposites.

3.1.4. SEM and EDS analysis

The morphologies of IM, $BaFe_{12}O_{19}$ nanoparticles, and $BaFe_{12}O_{19}$ @IM nanocomposite were investigated by SEM analysis and shown in Figure 7(a), (b) and (c), respectively. IM micrographs indicate an amorphous polymeric structure (Figure 7(a)). The SEM micrograph of the $BaFe_{12}O_1$ nanoparticles (Figure 7(b)) illustrates a uniform lace-like morphology made up of hexagonal polycrystalline nanoparticles with an average size of around 70 nm. The SEM image related to the $BaFe_{12}O_{19}$ @IM nanocomposites shown in Figure 7(c) shows well that the IM surface was practically uniformly covered by hexagonal $BaFe_{12}O_{19}$ @IM nanoparticles. The average size of $BaFe_{12}O_{19}$ @IM

nanocomposites is around 85 nm. The EDS analysis of $BaFe_{12}O_{19}$ @IM divulges the presence of carbon, sulfur, oxygen, barium, and iron in the structure of this material (Figure 7(d)).

3.1.5. Energy dispersive X-ray (XRD)

Figure 8 shows the X-ray diffraction patterns of the bare BaFe₁₂O₁₉, BaFe₁₂O₁₉@IM, and IM. The XRD pattern of BaFe₁₂O₁₉ displayed distinct diffraction peaks at 2θ values = 29.92, 31.68, 33.54, 34.84, 36.54, 54.61, 56.18, and 62.84, which are related to the (110), (107), (114), (203), (217), (2011), and (220)crystal planes of $BaFe_{12}O_{19}$ based on the [01-072-0738] standard chart. The amorphous polymeric structure of the IM polysaccharide can be highlighted by a typical broad peak at 20. Also, it is clearly obvious that the crystal structure of the barium hexaferrite has been preserved after treatment by IM without any significant changes in its XRD pattern. So, the main structural peaks of the BaFe₁₂O₁₉@IM nanocomposite and barium hexaferrite are the same. IM peaks exhibited low intensity in the BaFe₁₂O₁₉ @IM nanocomposite pattern relating to the robust intensity of the barium hexaferrite crystal phases, the weak peak intensity of the IM amorphous polymeric structure on the one hand and high mass fraction of the $BaFe_{12}O_{19}$ nanoparticles to the polysaccharide on the other.

3.1.6. Brunauer-Emmett-Teller (BET) analysis

The surface area, total pore volume, and average pore diameter of the $BaFe_{12}O_{19}$ @IM were analyzed by N_2 adsorption-desorption analysis and found to be 17.65 m²/g, 0.032 cm³/g, 7.29 nm respectively. As shown in Figure 9, $BaFe_{12}O_{19}$ @IM have type IV isotherms indicating mesoporous structures of the sample.

3.2. The antimicrobial activity of $BaFe_{12}O_{19}@IM$

The antibacterial activity of $BaFe_{12}O_{19}$ @IM was investigated against two bacterial strains: *E. coli* and *S. aureus*. The plate of Mueller-Hinton (MH) supplemented with tween 80 surfactants (final concentration of 0.05% v/v) using an agar medium was applied. Suspensions of each bacterium were accumulated to obtain in the vicinity of 108 colony forming units (cfu) per ml for agar plating. For this purpose, 50 mg of the sample were maintained at 4°C for 2 h, and then incubated overnight at 37°C. Clear inhibition zones around the discs indicated the presence of antimicrobial



Figure 7. SEM images of (a) IM, (b) bare $BaFe_{12}O_{19}$ nanoparticles, (c) $BaFe_{12}O_{19}@IM$ nanocomposite, and (d) EDS analysis of $BaFe_{12}O_{19}@IM$ nanocomposite.



Figure 8. X-ray diffraction patterns of the bare $BaFe_{12}O_{19}$, IM, and $BaFe_{12}O_{19}@IM$.



Figure 9. BET curve of BaFe₁₂O₁₉@IM.



Figure 10. Antibacterial activity of $BaFe_{12}O_{19}@IM$ against *S. aureus*.

activity. The diameter of the clear inhibition zone around the BaFe₁₂O₁₉ @IM disc is 10 mm for *S. aureus*, whereas no obvious growth was detected for *E. coli*. (Figure 10).

3.3. Catalytic activity of BaFe₁₂O₁₉@IM

The catalytic activity of the heterogeneous BaFe₁₂O₁₉@IM nanocomposite for the preparation of benzopyranopyrimidines was investigated. For this purpose, the pseudo four-component condensation of salicylaldehyde, and reaction malononitrile morpholine as a model reaction was studied. Some relevant results of these reactions are summarized in Table 2. In the beginning, the reaction was undertaken under catalyst-free conditions and there was not any progress in this situation (Table 2, entry 13). In the presence of $BaFe_{12}O_{19}$ MNPs and IM, the reaction proceeded with 30% and 60% yields, respectively (Table 2, entries 5, 6). However, the modified catalyst BaFe₁₂O₁₉@IM nanocomposite increased the yield of reaction up to 95% (Table 2, entry 2) due to the

Entry	Catalyst (mg)	Solvent	$\begin{array}{c} \mathbf{Yield} \\ (\%)^{\mathrm{b}} \end{array}$
1	BaFe ₁₂ O ₁₉ @IM (20 mg)	EtOH	95
2	$BaFe_{12}O_{19}@IM (15 mg)$	EtOH	95
3	$BaFe_{12}O_{19}@IM (10 mg)$	EtOH	90
4	$BaFe_{12}O_{19}@IM (5 mg)$	EtOH	80
5	$BaFe_{12}O_{19} (15 mol\%)$	EtOH	30
6	IM (15 mg)	EtOH	60
7	$BaFe_{12}O_{19}@IM (15 mg)$	$\mathrm{H}_{2}\mathrm{O}$	70
8	$BaFe_{12}O_{19}@IM (15 mg)$	MeOH	75
9	$BaFe_{12}O_{19}@IM (15 mg)$	$\mathrm{CH}_{2}\mathrm{Cl}_{2}$	35
10	$BaFe_{12}O_{19}@IM (15 mg)$	Toluene	40
11	$BaFe_{12}O_{19}@IM (15 mg)$	$\mathrm{CH}_3\mathrm{CN}$	75
12	$BaFe_{12}O_{19}@IM (15 mg)$		25
13	No catalyst	EtOH	Nil

^a: Reaction conditions: salicylaldehyde (2 mmol), morpholine (1 mmol), malononitrile (1 mmol), catalyst, solvent (4 mL), room temperature.

^b: Isolated yields.

synergistic effects of BaFe₁₂O₁₉ and IM. To determine the efficiency of the catalyst concentration, some experiments were probed in the presence of different amounts of the catalyst (5, 10, 15, 20 mg), giving benzopyranopyrimidines with 80%, 90%, 95% and 95% isolated yields. Thus, the best yield is accessible in the presence of just a 15 mg catalyst, and using more amounts of the catalyst did not increase the result to a considerable level (Table 2, entries 1-4). The optimal amount of BaFe₁₂O₁₉ @IM catalyst (15 mg) has been examined in various solvents. Under solvent-free conditions, a trace of the product was produced (Table 2, entry 12). Satisfactory results have been obtained with polar protic solvents such as H_2O , MeOH, and EtOH (Table 2, entries 2, 7, 8), rather than nonpolar or polar aprotic solvents such as CH_2Cl_2 , Toluene, CH_3CN (Table 2, entries 9, 10, 11).

Various derivatives of salicylaldehyde and secondary amines were perused under optimized reaction conditions, and the results are shown in (Table 3). The vast range of salicylaldehyde substitution can produce a reaction from electron-withdrawing groups like halogens up to electron-donating substituents such as the methoxy group. All products are known compounds, almost all the reactions worked quite well, and the desired products were obtained in good to high yields.

A comparison between the presented protocol in this study and other reported protocols for the same model reaction has been reported in Table 4. According to these results, the yield of benzopyranopyrimidine

Table 2. Optimization of the $BaFe_{12}O_{19}$ @IM nanocomposite for synthesis of the benzopyranopyrimidine derivatives $(3j)^{a}$.

	2 mmol la-g	$- \begin{array}{c} CN \\ CN \\ CN \\ 1 \text{ mmol} \\ 2 \\ 3a-e \end{array}$	BaFe ₁₂ O ₁₉ @IM Ethanol, r.t	R ² N ⁻ R ¹ N Ma-p X	OH
Entry	Aldehyde	Amine	Product	${f Yields^b}\ (\%)$	M.p. (°C) Found/Ref.
1	O H OH	Morpholine		95	197-199/196-197 [40]
2	O H OH	Piperidine	N N N N O H 4b	95	167-179/168-170 [32]
3	O H OH	Dimethylamine	N N N N N O H Ac	90	180-183/177-179 [32]
4	O H OH	Pyrrolidine	$ \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ $	80	235-237/235-237 [34]
5	Br H OH	Morpholine	Br H H H H H H H H H H H H H H H H H H H	83	200/194-196 [43]
6	O H OH OMe	Morpholine		85	212-215/210-220 [44]

Table 3. The pseudo four-component preparation of benzopyranopyrimidines catalyzed by $BaFe_{12}O_{19}@IM$ nanocomposite^a.

^a: Reaction conditions: Salicylaldehyde derivatives (2 mmol), amines (1 mmol), malononitrile (1 mmol), BaFe₁₂O₁₉@IM (0.015 g), solvent (4 mL), room temperature;

^b: Isolated yields.

	(continued).			_ 2 _ 1	
	X H H	CN R4RaNHa	BaFe ₁₂ O ₁₉ @IM	$\mathbb{R}^{2}_{N}\mathbb{R}^{1}$	ОН
	OH 2 mmol 1a-g	CN 1 mmol 1 mmol 2 3a-e	Ethanol, r.t	O N K	
Entry	Aldehyde	Amine	Product	$egin{array}{c} \mathbf{Yields}^{\mathrm{b}} \ (\%) \end{array}$	M.p. (°C) Found/Ref.
7	Cl H OH	Morpholine	$C^{I} \rightarrow C^{I} \rightarrow C^{I}$	90	249-251/247-250 [45]
8	MeO OH	Diethylamine	$Br \xrightarrow{N} OH$	75	175-178/178-179 [46]
9	Br H OH	Piperidine	Br H H H H H	80	222-225/226 [43]
10	O H OH OMe	Piperidine		92	176-179/178-180 [44]
11	Cl H OH	pyrolidine		96	192-194/191-194 [47]
12	O H OH OMe	pyrolidine	$ \begin{array}{c} \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ $	95	157-159/158-160 [43]

Table 3. The pseudo four-component preparation of benzopyranopyrimidines catalyzed by $BaFe_{12}O_{19}@IM$ nanocomposite^a (continued).

^a: Reaction conditions: Salicylaldehyde derivatives (2 mmol), amines (1 mmol), malononitrile (1 mmol), BaFe₁₂O₁₉@IM (0.015 g), solvent (4 mL), room temperature;

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^b: Isolated yields.



Table 3. The pseudo four-component preparation of benzopyranopyrimidines catalyzed by $BaFe_{12}O_{19}$ @IM nanocomposite^a (continued).

^a: Reaction conditions: Salicylaldehyde derivatives (2 mmol), amines (1 mmol), malononitrile (1 mmol), BaFe₁₂O₁₉@IM (0.015 g), solvent (4 mL), room temperature;

^b: Isolated yields.

Table 4. Comparison of the activity of the catalysts for the synthesis of benzopyranopyrimidine (4a) with some of the other catalysts reported in the literature.

Entry	Catalyst	Reaction condition	Reaction time (h)	${f Yield} \ (\%)$	Reference
1	$LiClO_4$	Ethanol, rt	24	84	[32]
2	Piperidine	$100^{\circ}\mathrm{C}$	11	78	[33]
3	Fe(II)-BTU-SNPs	Ethanol, rt	4	93	[34]
4	Tetrabromobenzene-1,3-disulfonamide	Ethanol, rt	24	90	[42]
5	$\rm Fe_3O_4/IRMOF-3/SO_3H$	Ethanol, rt	1	95	[36]
6	p-toluenesulfonic acid	Solvent-free, rt	1	75	[50]
7	$BaFe_{12}O_{19}@IM$	Ethanol, rt	1	95	This work

production in the present study was 95%, which is effectively better than the other projects with different catalysts. Also, in terms of reaction time, the production of benzopyranopyrimidine was 1 hour which is lower than the other projects. Totally, these two differences between present and prior results can confirm the efficiency of this project. The proposed mechanism for the formation of the benzopyranopyrimidines with the desired consequence of the catalytic activity of $BaFe_{12}O_{19}$ @IM nano-catalyst is shown in Figure 11. Referring to the suggested mechanism, Brønsted acidic sites of the catalyst, i.e. -OH and metal positively-charged metal ions (Fe³⁺, Ba²⁺), which facilitate the Knoevenagel



Figure 11. Proposed mechanism for the synthesis of benzopyranopyrimidines in the presence of $BaFe_{12}O_{19}$ @IM magnetic nanocomposites.

condensation reaction, might activate the carbonyl group of salicylic aldehyde toward nucleophilic attack of the malononitrile anion generated by Lewis basic sites of IM ($-OSO_3^-$) and (O^{2-}) of BaFe₁₂O₁₉ in the catalyst. On the other hand, these Lewis basic sites in the catalyst, by activating the phenolic O-H groups, trigger the Pinner reaction which leads to the intermediate (I). Following that, the intermediate (II) is produced by the attack of the amine on the cyano group. Ultimately, intermediate (II) reacts with another salicylic aldehyde, giving rise to the intermediate (III), followed by hydrogen transfer to yield the final benzopyranopyrimidine. In fact, the formation of the product results from the activation of the reactants by the brønsted basic and acidic catalyst sites.

4. Conclusions

In conclusion, a natural polymer supported biomagnetic material, $BaFe_{12}O_{19}$ @IM, has been prepared using a simple sol-gel method. The catalytic activity of this biomagnetic material, as a heterogeneous bifunc-

tional acid-base catalyst, has been investigated in the synthesis of benzopyranopyrimidine derivatives. As the results have shown, the separation of this biomagnetic catalyst is very easy thanks to the effective magnetic property of $BaFe_{12}O_{19}$. The bifunctional character of the catalyst due to the existence of sulfate and hydroxyl groups in sulfated polysaccharide, together with the greater surface accessibility of these functional groups, besides the magnetic characteristic of the catalyst, are the key factors for catalytic performance, easy separation, and recyclability of BaFe₁₂O₁₉@IM. Study on the antibacterial performance of BaFe₁₂O₁₉@IM nanocomposites on bacteria strains was evaluated. The results showed that the prepared nanocomposites possess antibacterial activity against Gram-positive S. aureus.

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